

Granule membranes play dice. The quantal nature of secretion

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To the memory of Bruno Bassan

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E. NITZANY, I. HAMMEL, I. MEILIJSON, *JTB 2010*

I. HAMMEL, I. MEILIJSON, *JRSI 2012*

I. HAMMEL, I. MEILIJSON, *JRSI 2013*

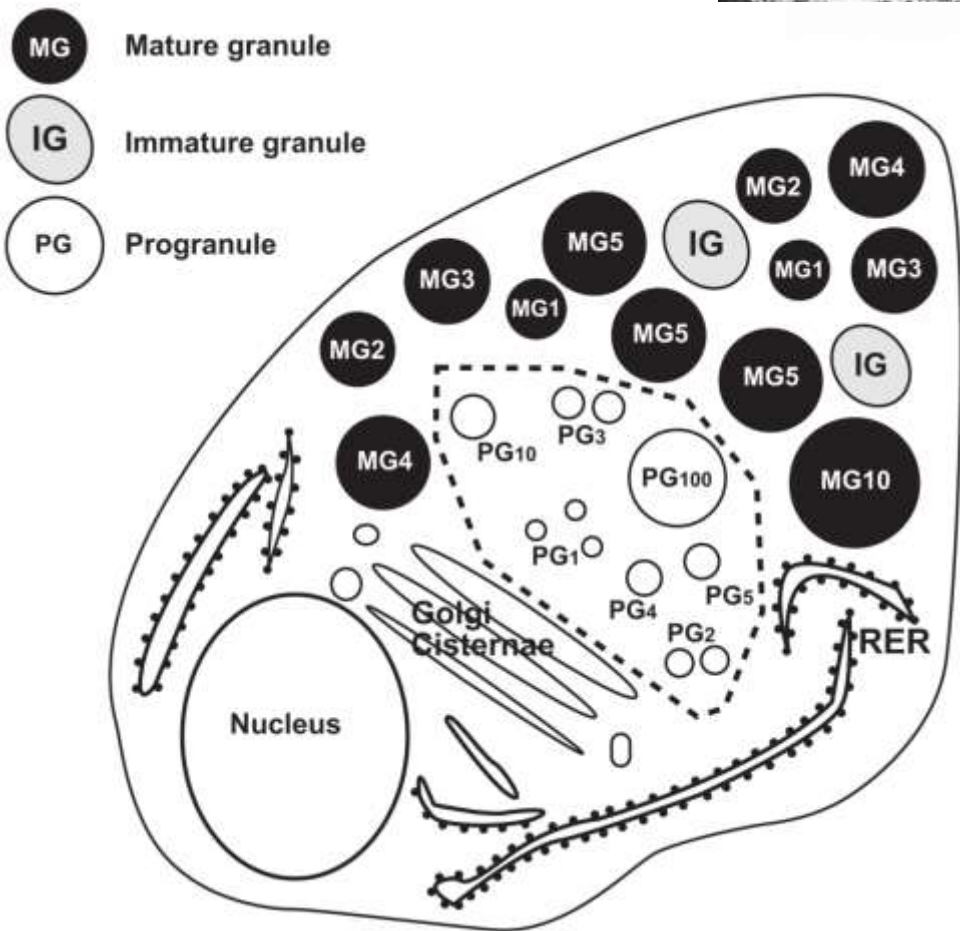
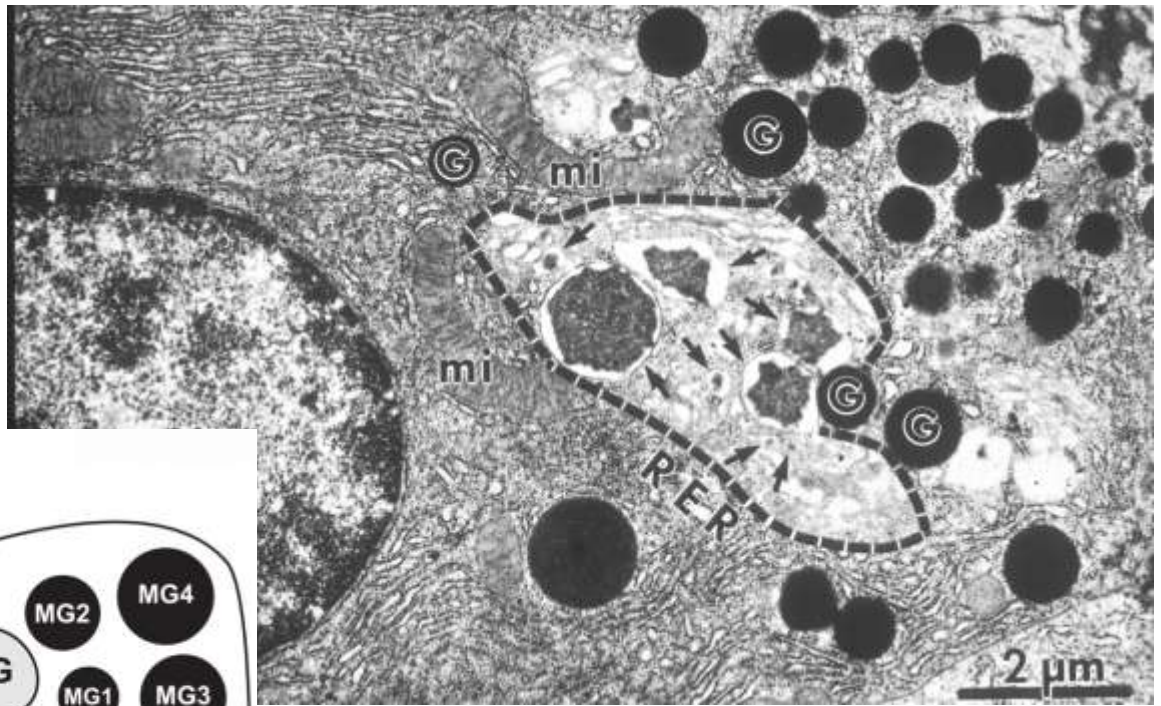
I. HAMMEL, M. KREPELOVA, I. MEILIJSON, *MRT 2014*

I. HAMMEL, I. MEILIJSON, *MI 2014*

I. HAMMEL, I. MEILIJSON, *NCB 2014*

I. HAMMEL, I. MEILIJSON, *D 2014*

D. AMIHAI, I. HAMMEL, I. MEILIJSON, J. TERKEL, *TO BE SUBMITTED*

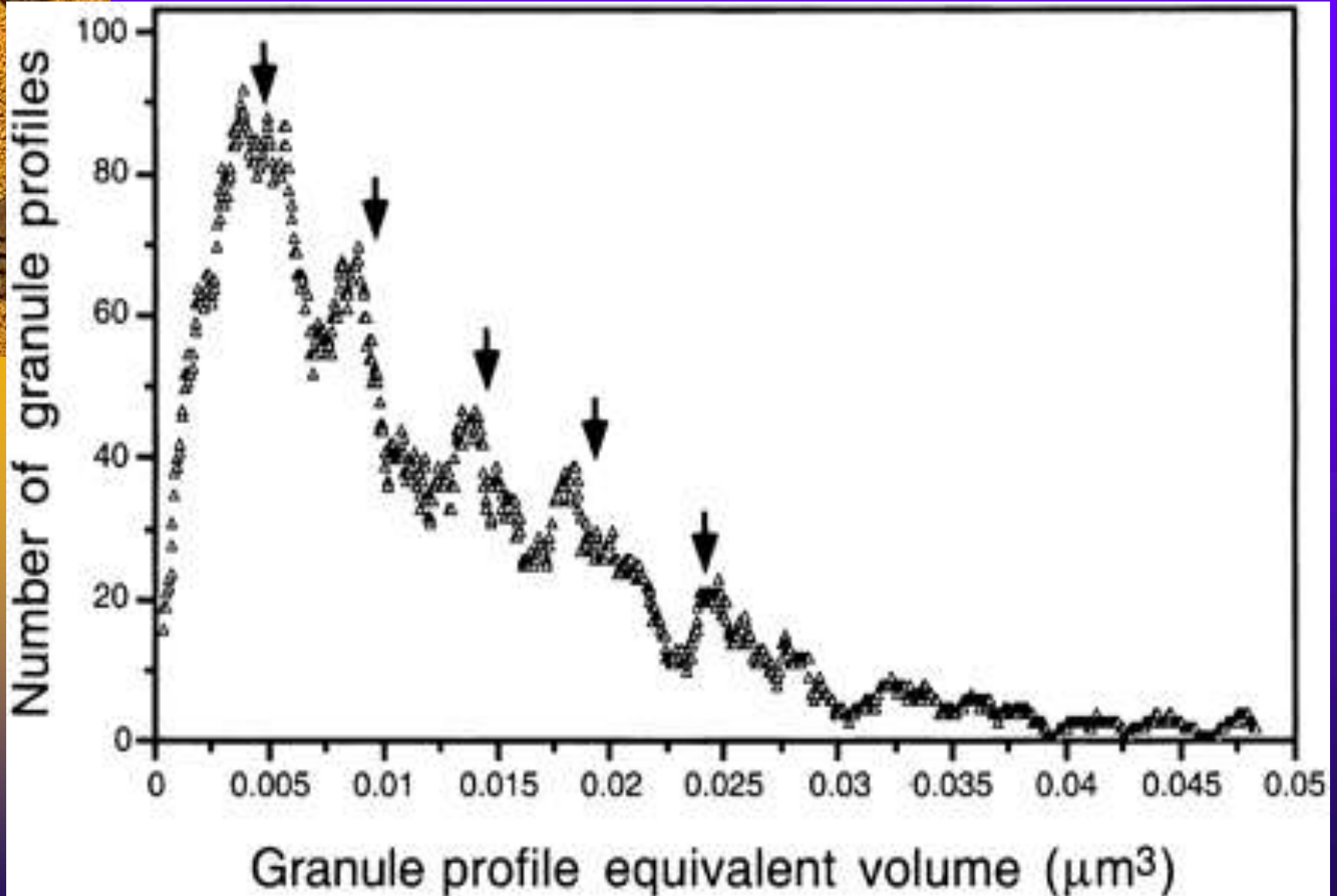


Pancreatic Acinar Cell

Two fundamental reasons for keeping granule inventory:

- 1. Uncertainty of demand:** Granule stock is maintained as a buffer to meet uncertainty in demand by the extracellular environment.
- 2. Lead time for production:** A source of supply during the lead time to produce granules of adaptive content.

Quantal nature of granular volume



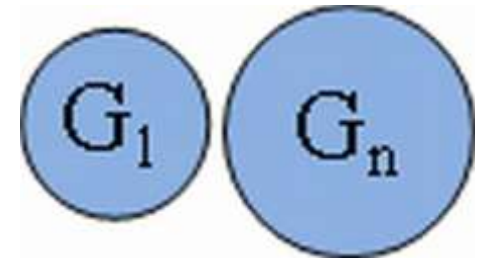


Most researchers hypothesize that granules stay as created and exit many-at-a-time

Bernard Katz (Nobel Prize 1970) group in England

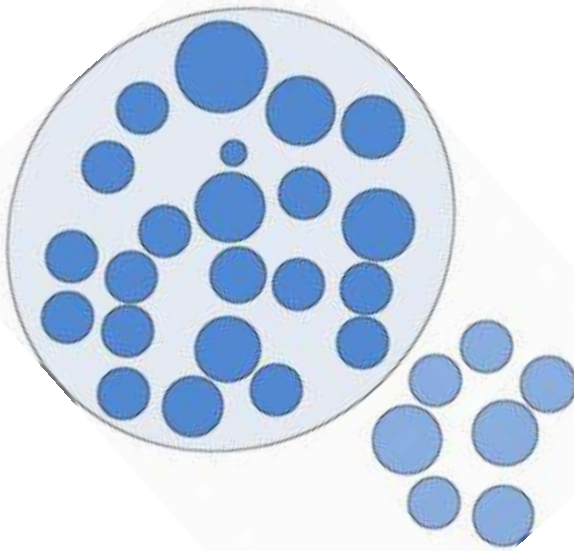
Cope & Williams 1992 review

Is *Quantal size* the synaptic content of a single vesicle out of a heterogeneous pool?



$$G_n = nG_1$$

Is *Quantal content* the number of homogeneous vesicles released in a single response of cell activation?



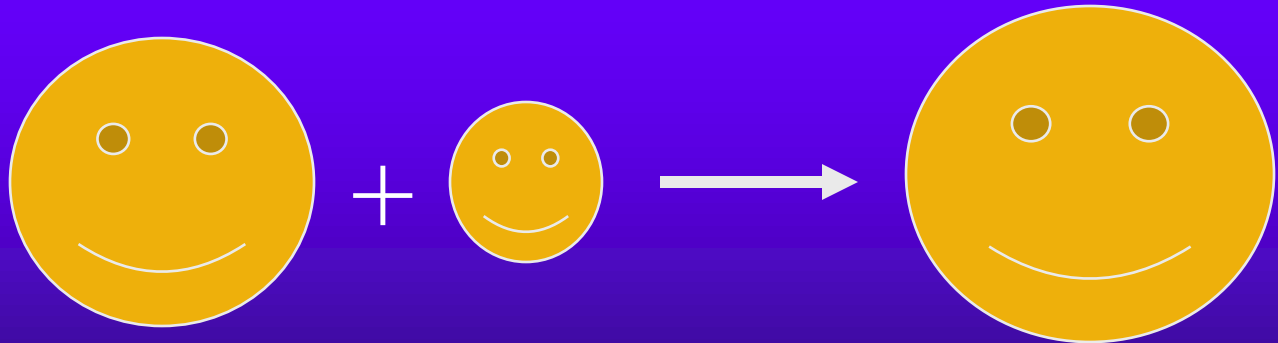
Granule Quantal Growth

addition of unit granule

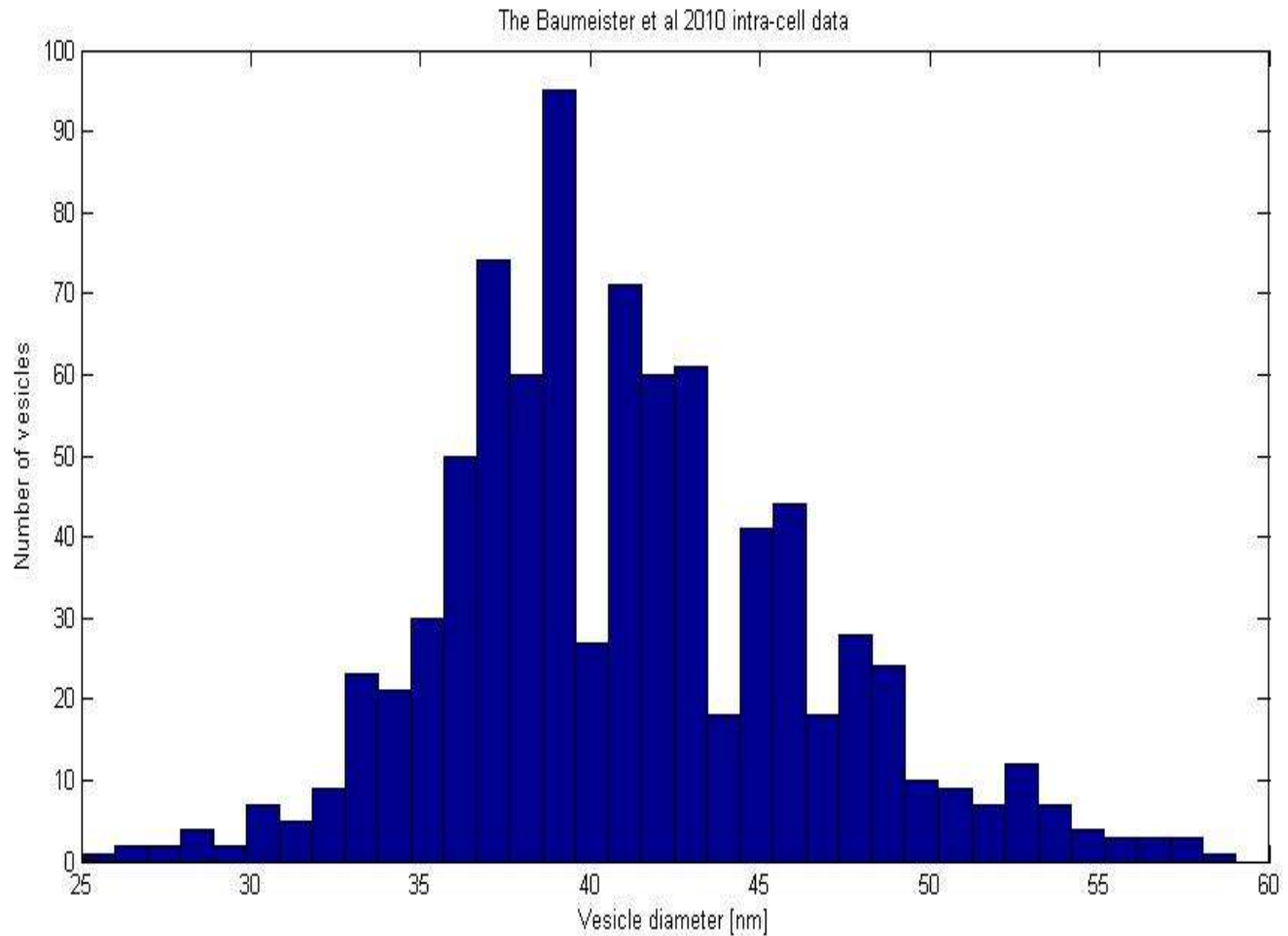


All agree on Granule-Membrane fusion

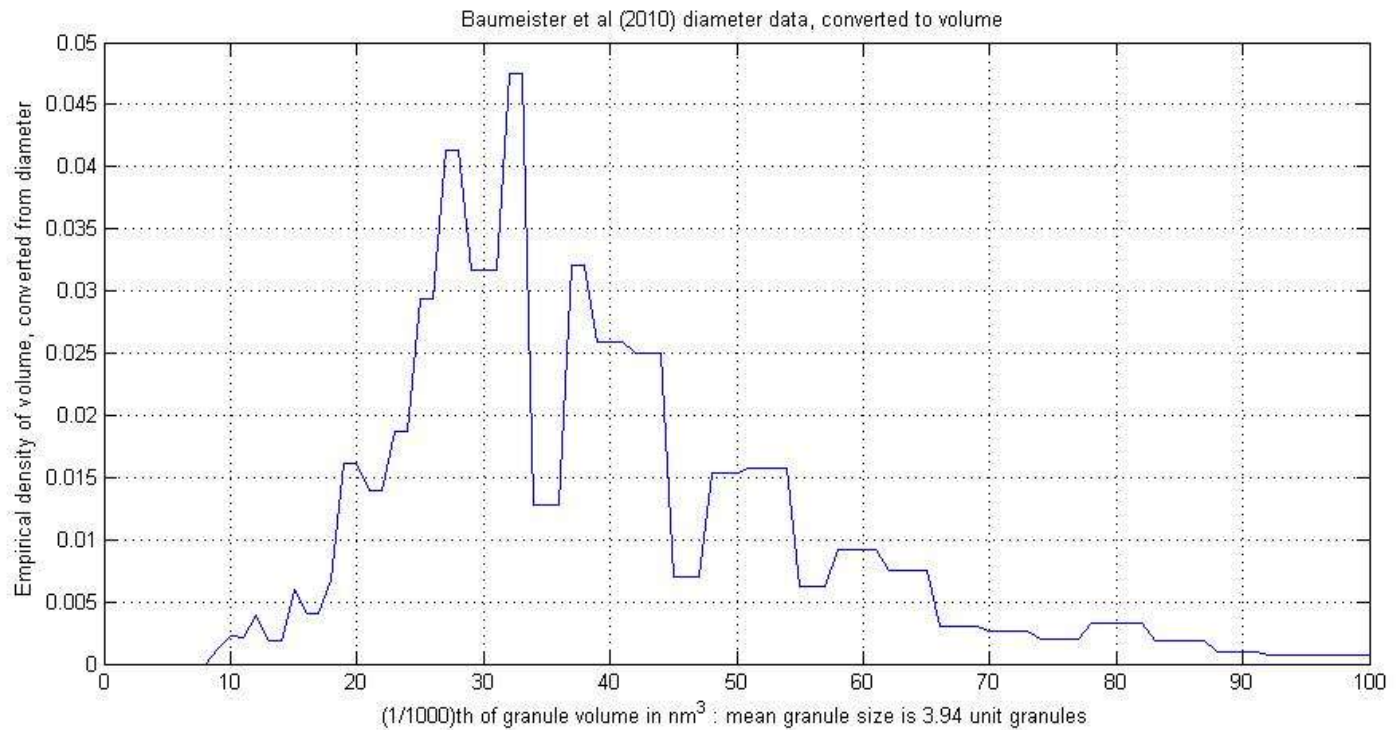
But not on Granule - Granule fusion



Baumeister et al 2010 diameter data



Histogram of volume data





Markov Stationary Model

Markov process: The distribution of Future given History is summarized by Present state.

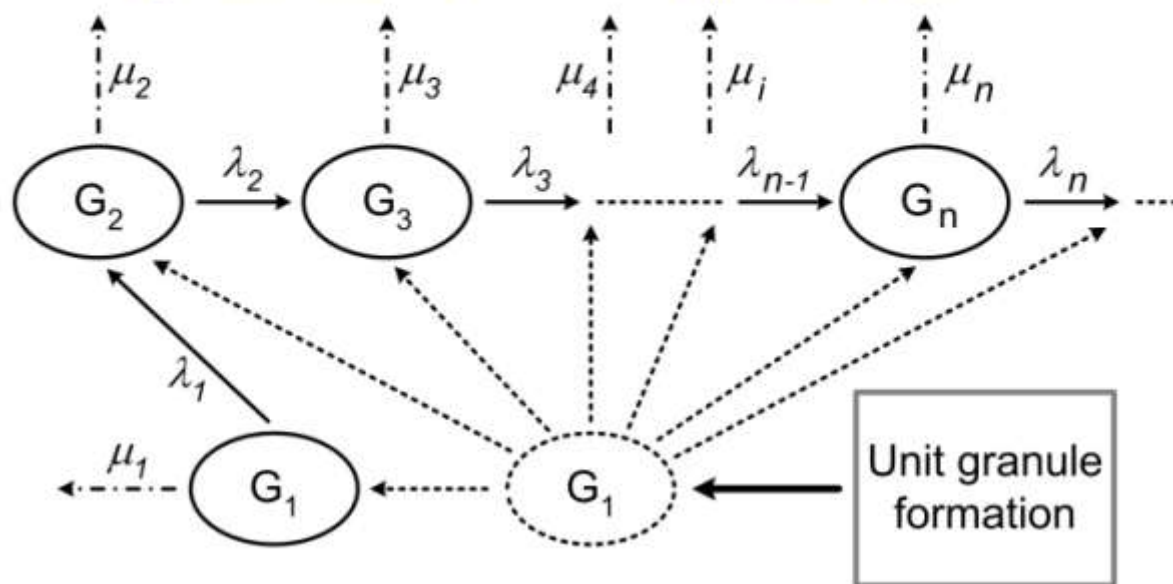
Granule Growth & Elimination : “state” is granule size



Probability of advancing from size n to size $n+1 = \frac{\lambda_n}{(\lambda_n + \mu_n)}$

$$\lambda_n = \lambda n^\beta = \lambda n^{-(2/3)(K_\beta - 1)}$$

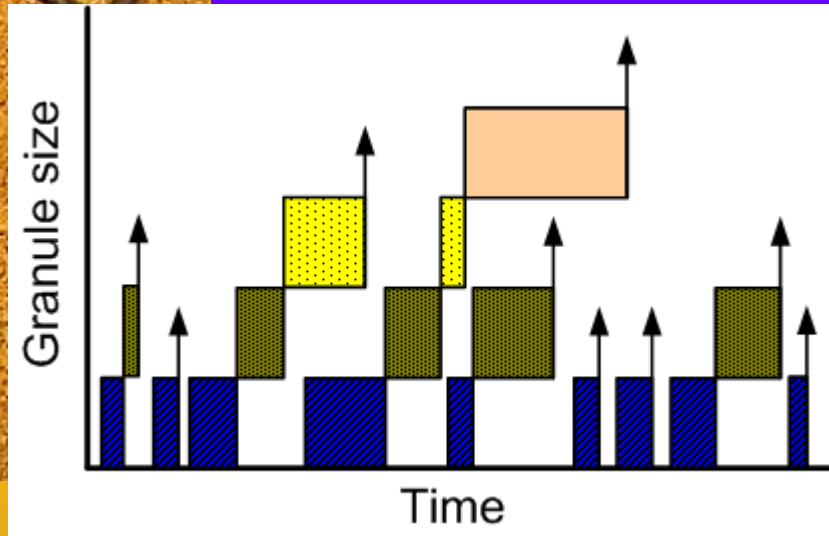
$$\mu_n = \mu n^\gamma = \mu n^{-(2/3)(K_\gamma - 1)}$$



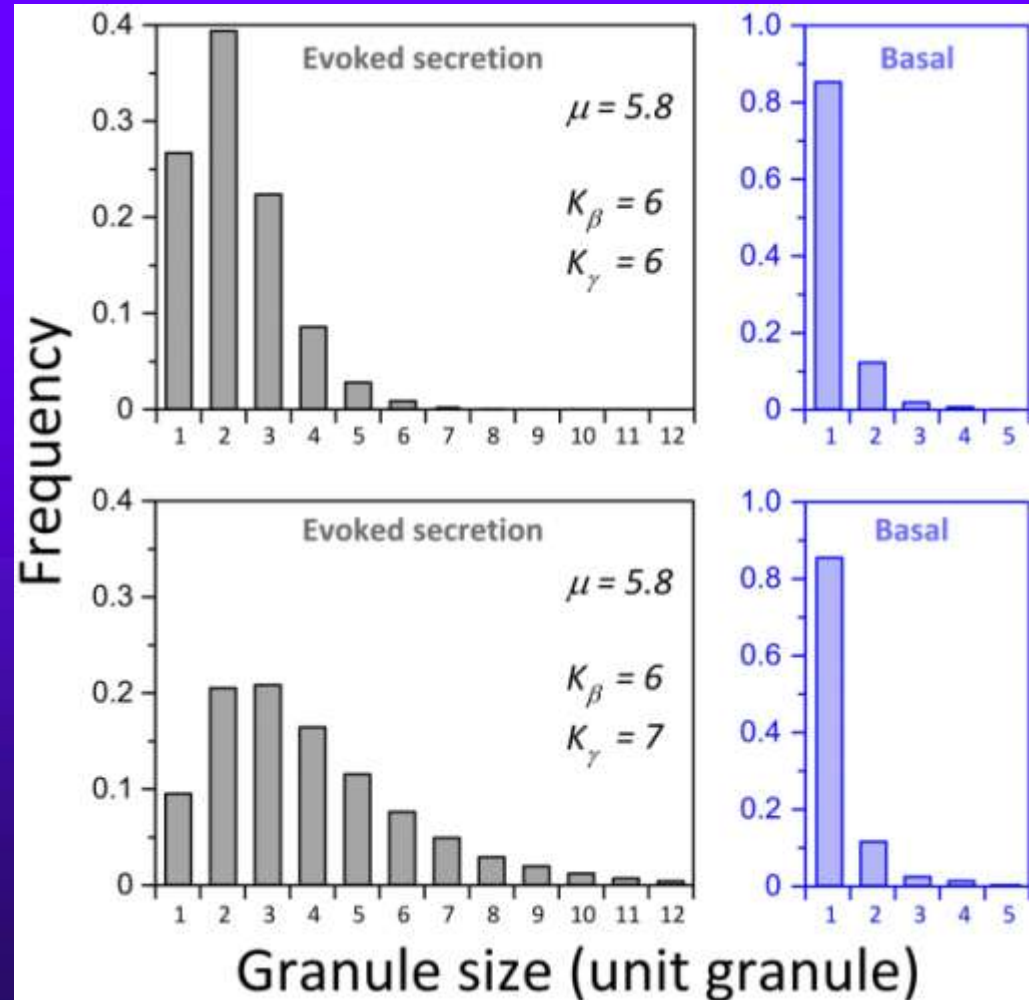
λ = addition
 μ = elimination

Granule steady-state model

Stationary distribution \neq Exit distribution



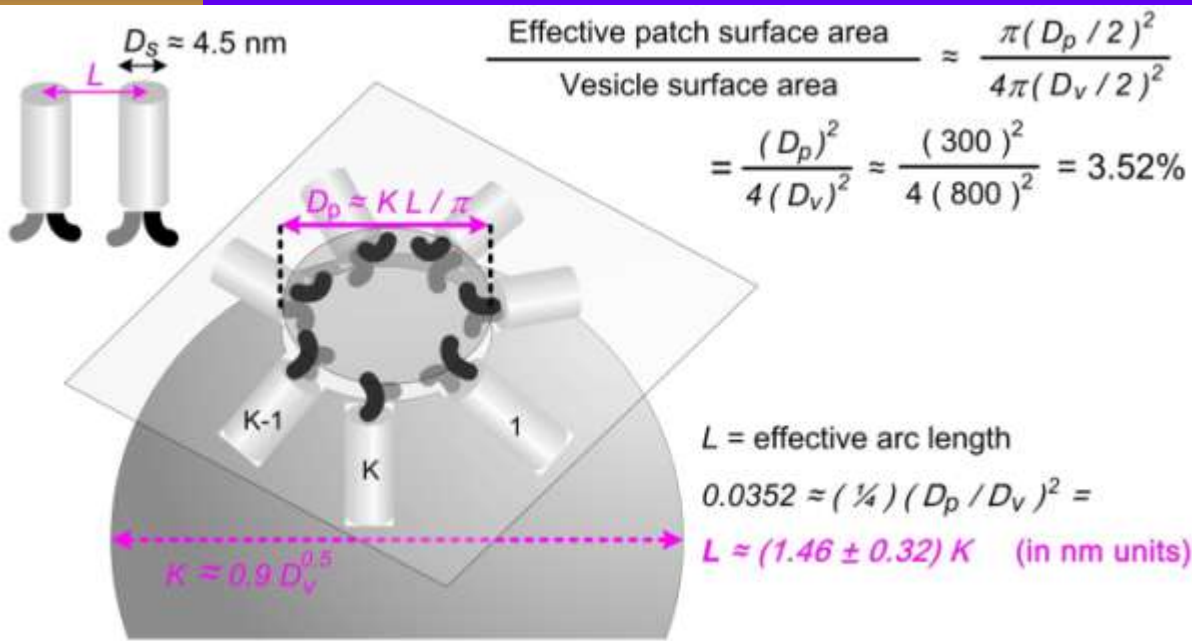
Ryan et al. (1997) used optical microscopy and fluorescent dyes to track the **spontaneous** and **evoked** vesicle release.



Three-parameter model

$$\mu/\lambda, \gamma, \beta$$

- ◆ Exit rate from n : $\mu * n^\gamma$
- ◆ Transition rate from n to $n+1$: $\lambda * n^\beta$
- ◆ Markov: independent exponentials, earliest wins
- ◆ Mean sojourn in n : $1/(\mu * n^\gamma + \lambda * n^\beta)$
- ◆ Probability of exit $\mu * n^\gamma / (\mu * n^\gamma + \lambda * n^\beta)$

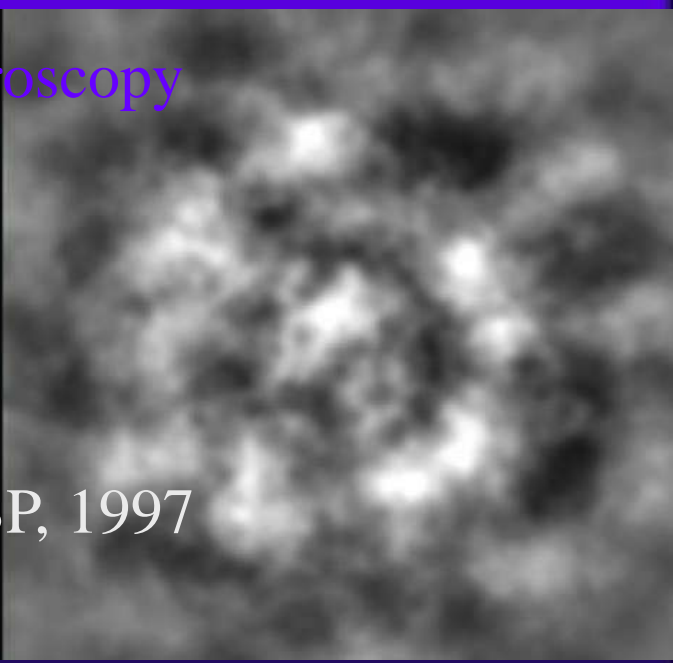


SNARE, ROSETTE, POROSOME

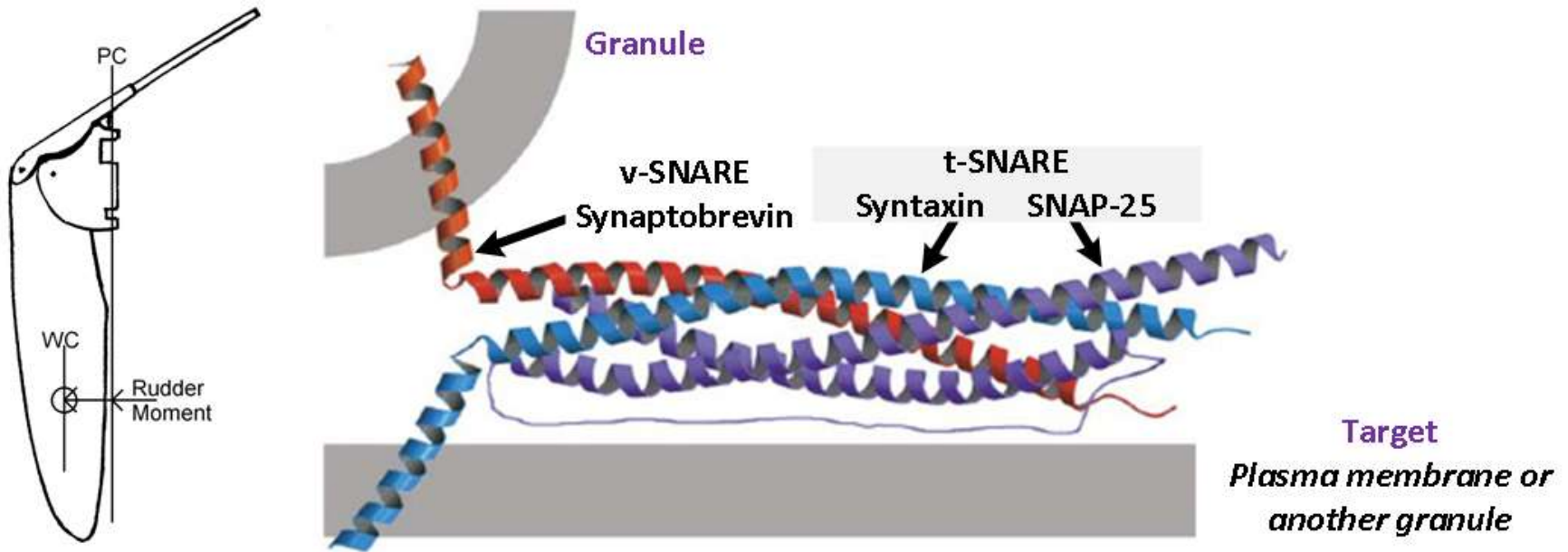


Atomic force microscopy

Porosome - Jena BP, 1997



The rudder: The SNARE SYSTEM



*Human subtlety will never devise an invention **more beautiful, more simple or more direct** than does nature because in her inventions nothing is lacking, and nothing is superfluous.*

—Leonardo da Vinci

http://www.brainyquote.com/quotes/authors/l/leonardo_da_vinci.html

Rationale for rate shape

SNARE: $\mu * n^\gamma$ and $\lambda * n^\beta$



N “rings” diffuse randomly on surface (area $n^{2/3}$) of granule, of which **K** must be close to each other and to **K** “hooks” in unit granule or membrane. Probability is of order

$(\text{const}/\text{area})^{K-1} = \text{const} * n^{-(2/3)(K-1)} = \mu * n^\gamma$ or $\lambda * n^\beta$

Similar to Hua & Scheller PNAS 2001



Steady-state condition: flow into $n+1$ equals flow out of $n+1$

$$\text{STAT}(n) * \lambda * n^{\beta} =$$

$$\text{STAT}(n+1) * (\mu * (n+1)^{\gamma} + \lambda * (n+1)^{\beta})$$

EXIT distribution

$$\text{EXIT}(n+) = \text{GROW}(1) * \\ \text{GROW}(2) * \dots * \text{GROW}(n-1)$$

and

$$\text{GROW}(m) = \lambda * m^\beta / (\mu * m^\gamma + \lambda * m^\beta)$$

$$\text{EXIT}(n) = \\ \text{EXIT}(n+) - \text{EXIT}((n+1)+)$$





The role of γ

- ◆ $\text{STAT}(n)/\text{EXIT}(n) = \exp\{-\gamma*\log(n)-b(\gamma)\}$
- ◆ **STAT** is exponential-type family with **EXIT** as case $\gamma=0$
- ◆ Perhaps evoked case needs **one** hook-ring pair!
- ◆ Reminder: $\gamma = -(2/3)(K-1)$

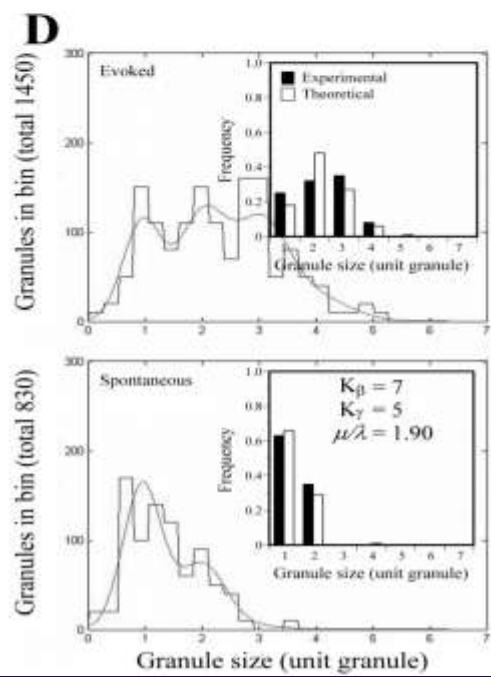
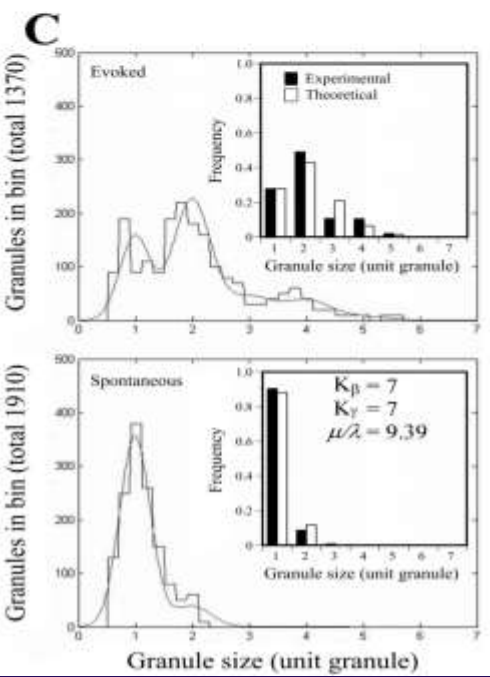
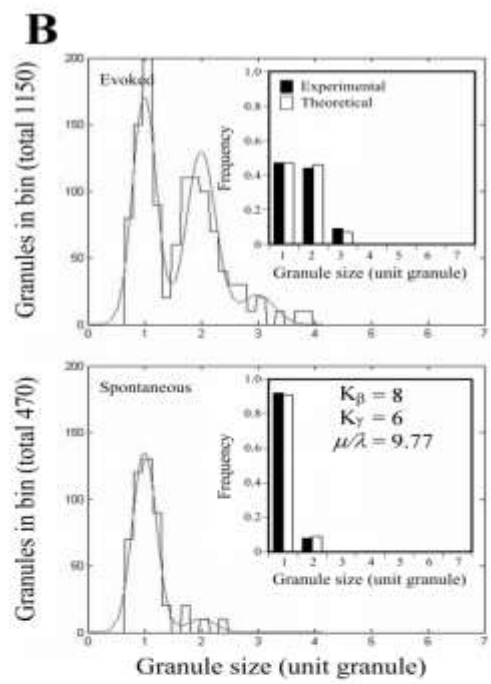
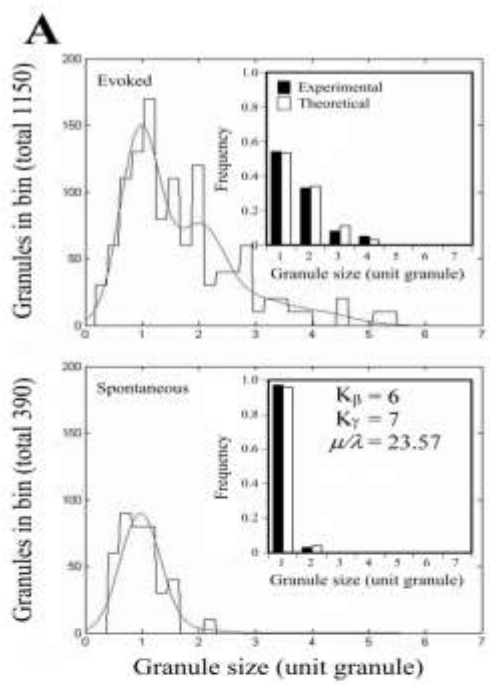


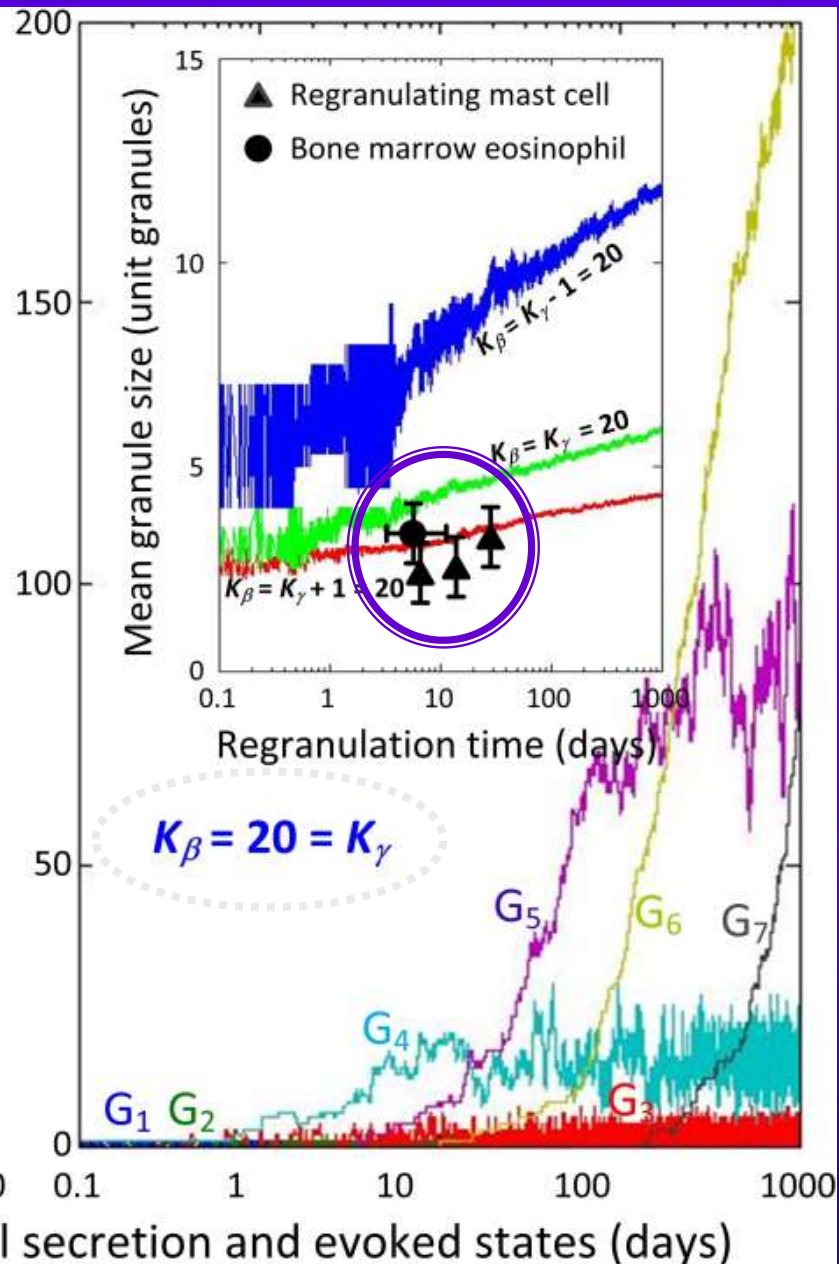
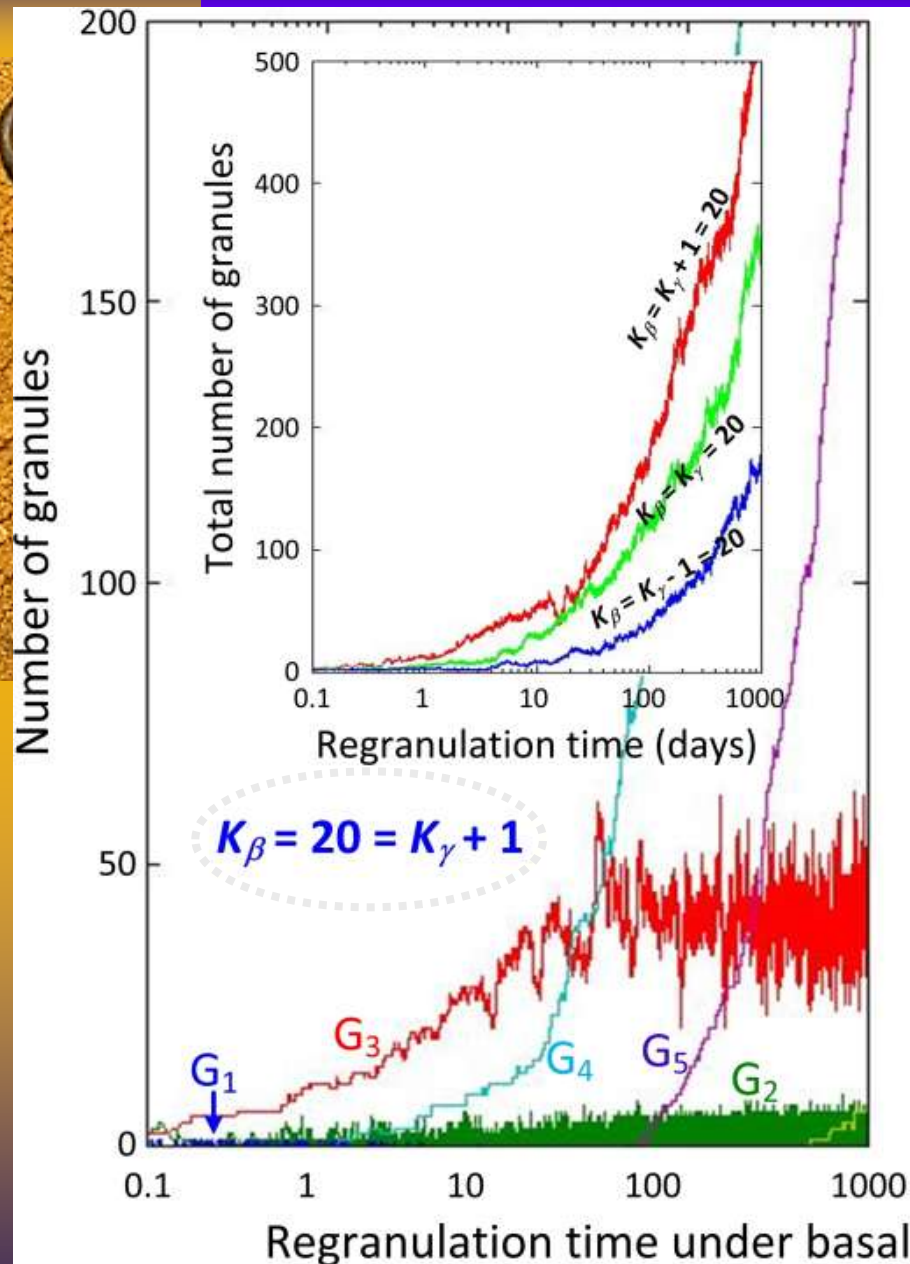
Statistical tools

- ◆ EM algorithm for Gaussian mixture with equally spaced means and variance
- ◆ MLE of μ/λ , γ , β based on evoked and spontaneous data
- ◆ Omnibus program for five parameters
- ◆ CUSUM detection of change-point
- ◆ Simulation study of transient behavior
- ◆ Euclidean Geometry considerations



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Detecting that spontaneous secretion turned evoked secretion

- ◆ CUSUM - Statistical tool for detecting change-point in distribution
- ◆ Performance measured by Kullback-Leibler divergence KLD
- ◆ CUSUM calculations may use common action potential monitoring.



CUSUM of Page (1953)

- ◆ Declare change when log likelihood
 - ◆ $S_n = \sum \log g(X_i)/f(X_i)$ exhibits draw-up
 - ◆ $S_n - \min_{m \leq n} S_m$
 - ◆ of at least a pre-assigned size.
-
- ◆ $E_g[\log g(X)/f(X)] > 0$ is $KLD(g, f)$
 - ◆ $E_f[\log g(X)/f(X)] < 0$ is $KLD(f, g)$



SUMMARY, FORMULA

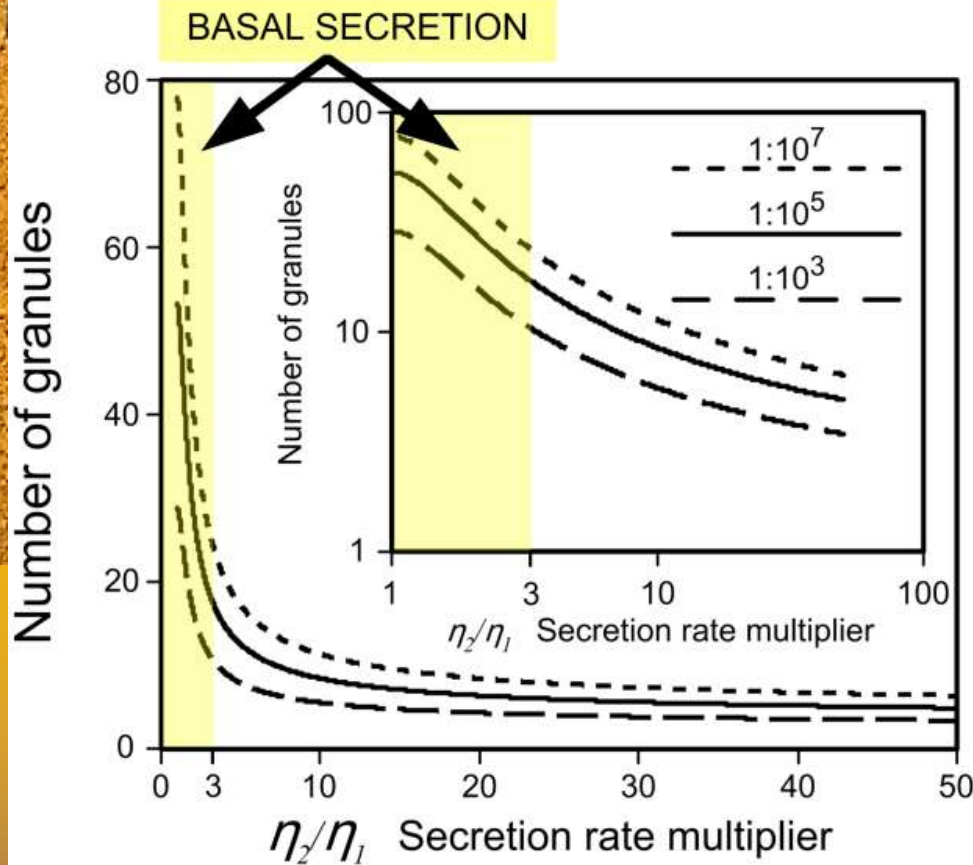
- ◆ For log likelihoods $\rho=1$.
- ◆ *Putting all together, Mean Granules to Detection is*

$$\ln[MGFA] + \ln[KLD(E,S) + \eta_2/\eta_1 - 1 - \ln(\eta_2/\eta_1)] \quad \blacklozenge$$

----- \blacklozenge

- ◆ $KLD(S,E) + \eta_1/\eta_2 - 1 - \ln(\eta_1/\eta_2)$

One word = 1 bit = 8-10 vesicles



If spontaneous secretion is steady at (the common rate of neurons) 1Hz, a false recognition alarm will be declared once every 15 minutes (10^3), once a day (10^5) or every four months (10^7).



Cellular communication: do we really need granule growth?

If the rate of secretion is increased

- ◆ by a small factor (≈ 2), granule size distribution plays a critical role
- ◆ by significant bursts (rate of increase = $\eta_2 / \eta_1 > 10$) the role of granule size distribution is minor.

The G&E model suggests that *granule polymerization has an advantage for the information gain it achieves under limited exocytosis of a small number of granules.*